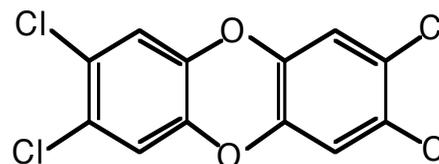
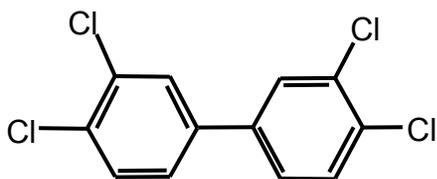


CHE Partnership call - April 2017

Gene Editing: Where Genomic Technologies Meet Environmental Health

CRISPR ways to understand gene-environment interactions



Mark Hahn

with Neel Aluru, Sibel Karchner,
and many other colleagues and collaborators



Woods Hole Oceanographic Institution, Woods Hole, MA

Boston University Superfund Basic Research Program

<http://www.busrp.org>



Boston University
Superfund Research Program



Outline

- Gene-Environment Interactions
- Chemical-adapted populations of fish
- Population genomics reveals critical role of aryl hydrocarbon receptor (AHR) signaling pathway
- CRISPR-Cas9 gene targeting
- Targeting AHR pathway genes in a non-model organism
- Significance

Gene-Environment Interactions

- Genes control response to chemicals
 - pollutants, drugs, nutrients
 - how sensitive?
 - metabolism and excretion

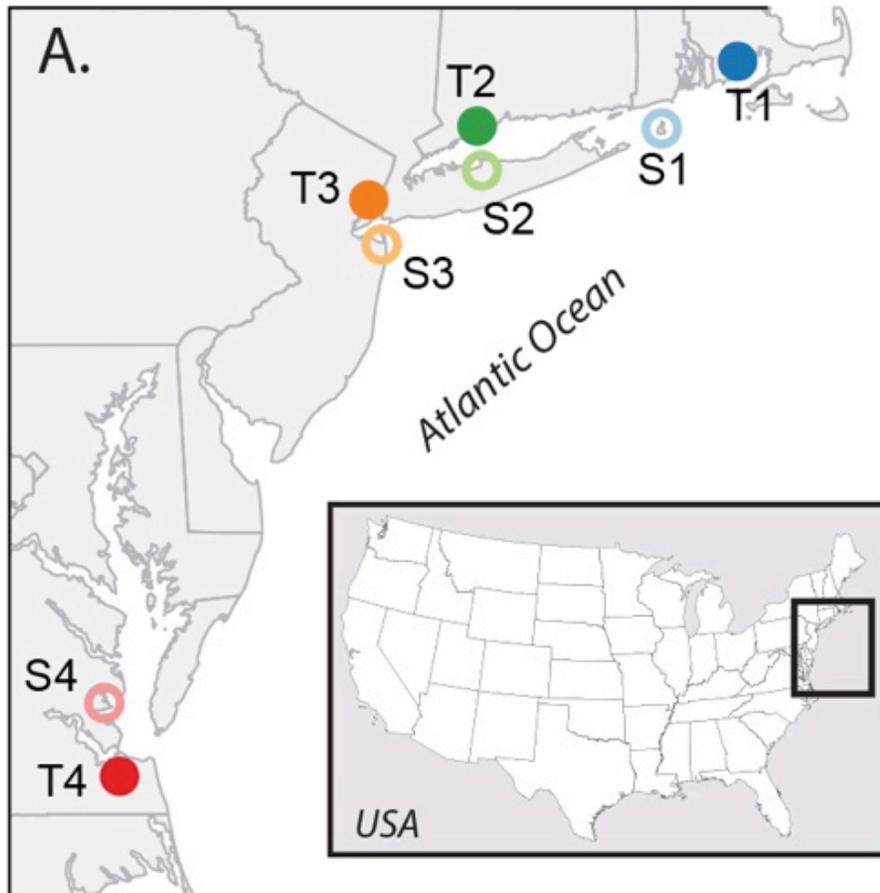
- Chemicals affect genes
 - altered gene expression
 - genetic damage
 - selection for genetic variants

Ecological Impacts at Superfund Sites

What is the impact of long-term (multi-generational) exposure to contaminants at Superfund sites?

- Effects on populations; multiple generations.
- Adaptation to rapidly changing environment?
- Mechanisms of adaptation?
 - Molecular basis?
 - Independent or shared among sites?
- Costs / Trade-offs

Tolerance (resistance) to aromatic hydrocarbons in four populations of Atlantic killifish *Fundulus heteroclitus*

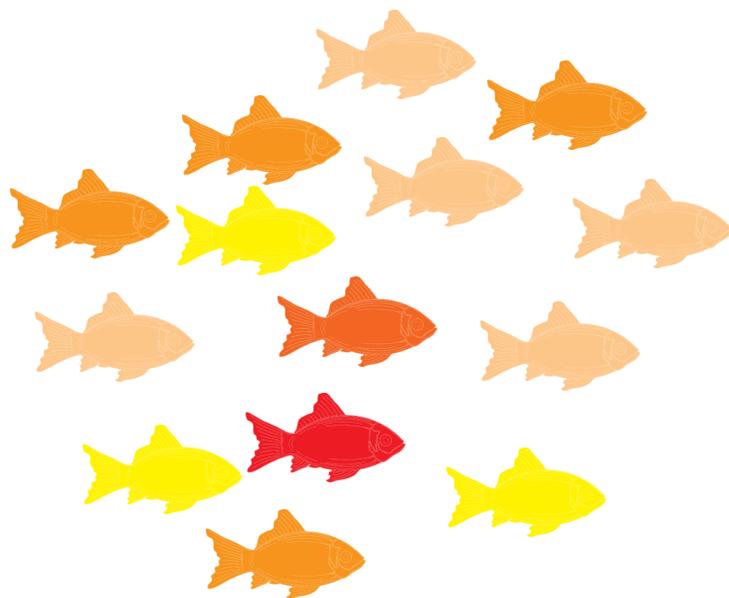
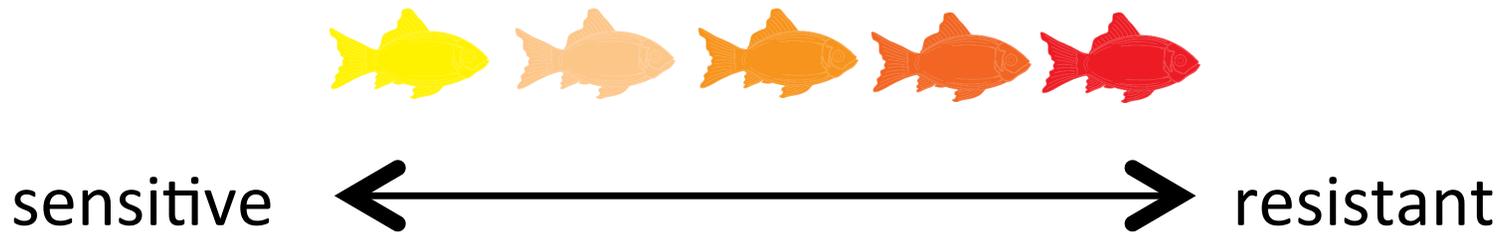


T=Tolerant S=Sensitive

- New Bedford Harbor, MA PCBs
- Bridgeport, CT PCBs, PAHs
- Newark, NJ Chlorinated dioxins
- Elizabeth River, VA PAHs

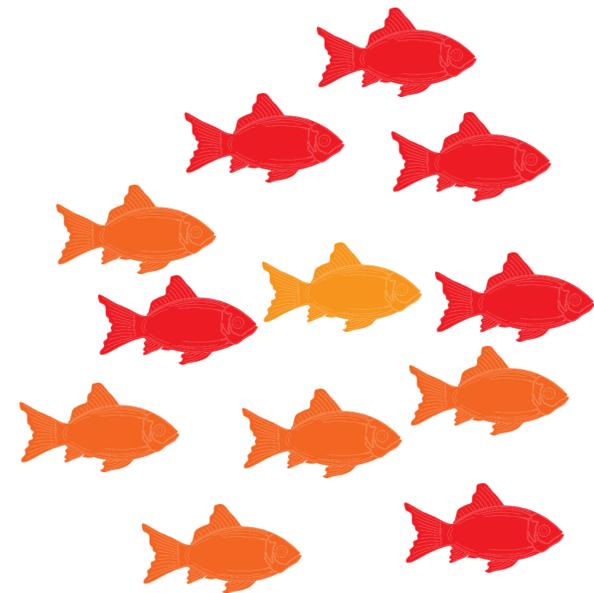
Reid *et al.* 2016 *Science* 354: 1305

Evolution of Resistance by Natural Selection



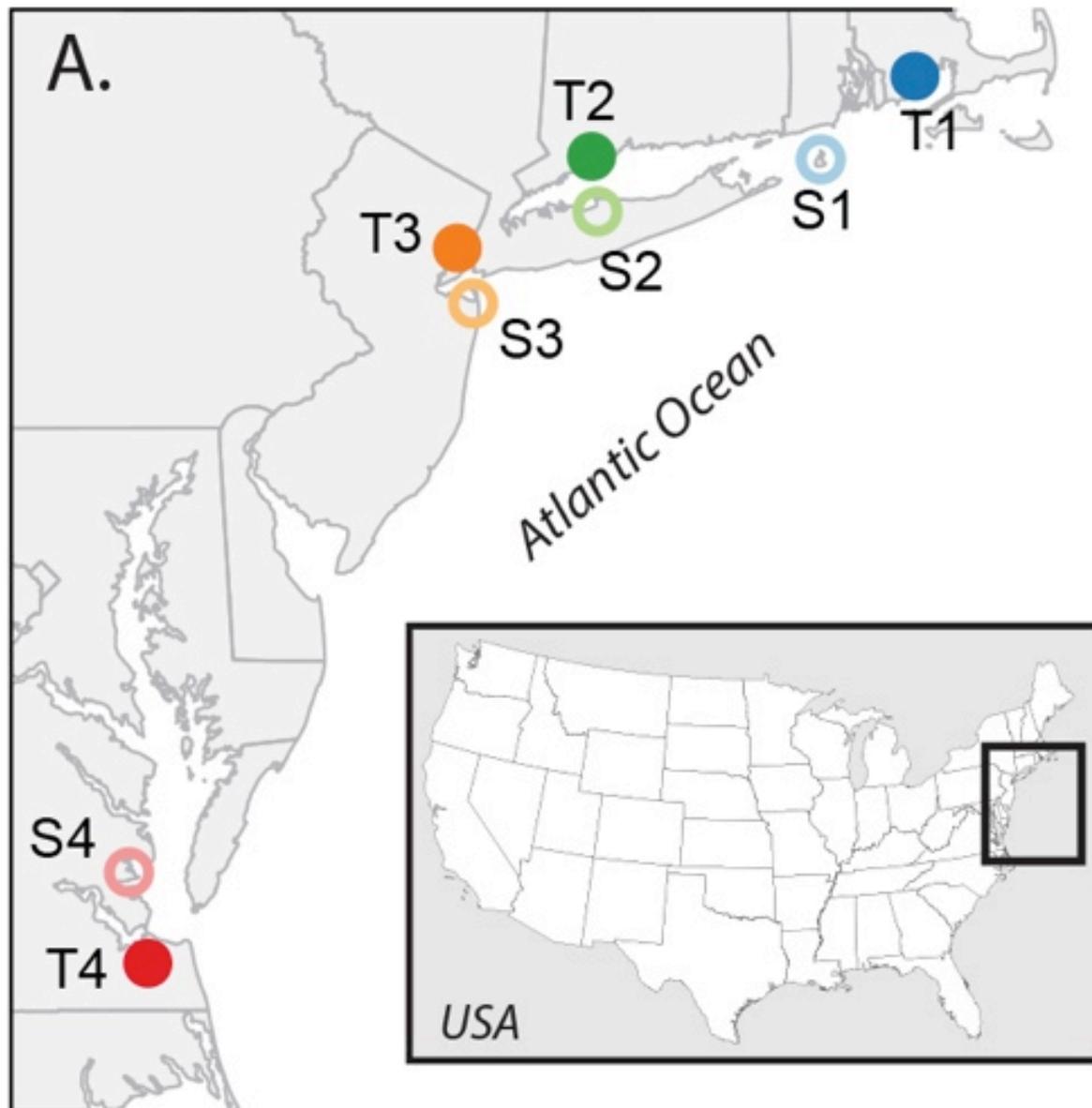
original population
(1940)

PCBs
→ →
multiple
generations



current population
(2017)

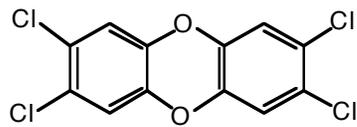
Population genomic study



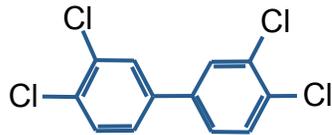
- Eight populations
- Four T-S pairs
- 48 fish per population
- Transcriptomics (gene expression)
- Genome sequencing
- Identification of genes under selection

Reid *et al.* 2016 *Science* 354: 1305

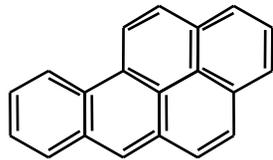
Aryl Hydrocarbon Receptor (AHR) Pathway



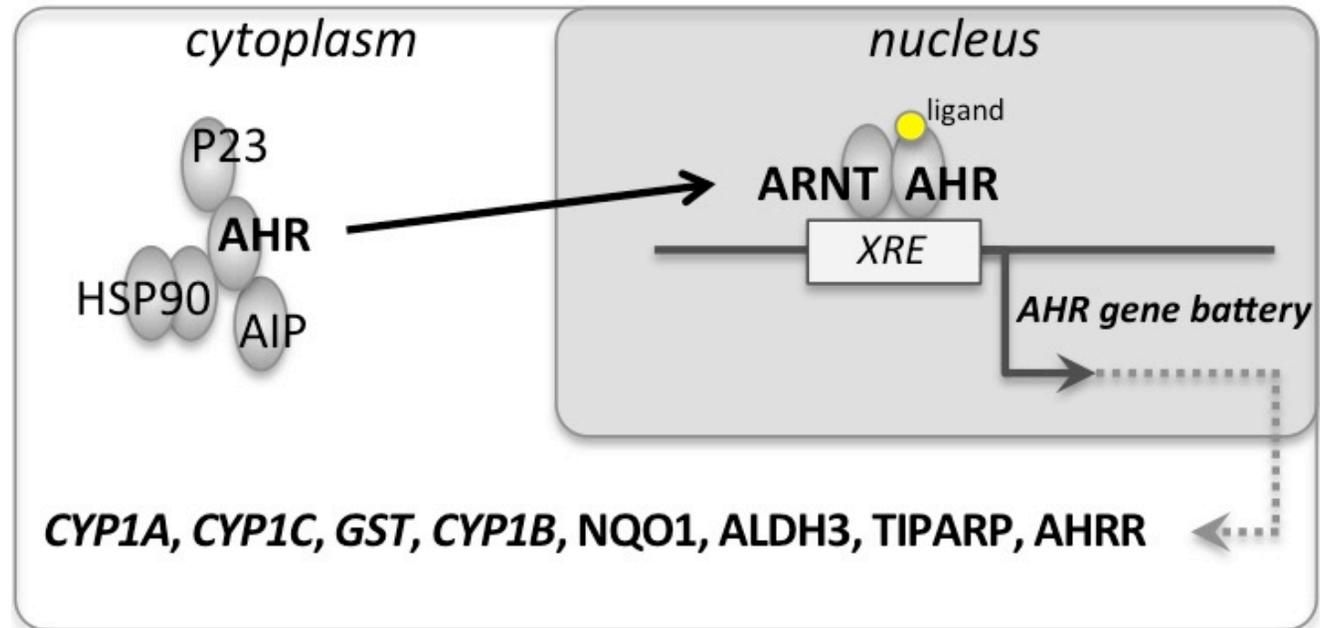
TCDD (dioxin)



PCB

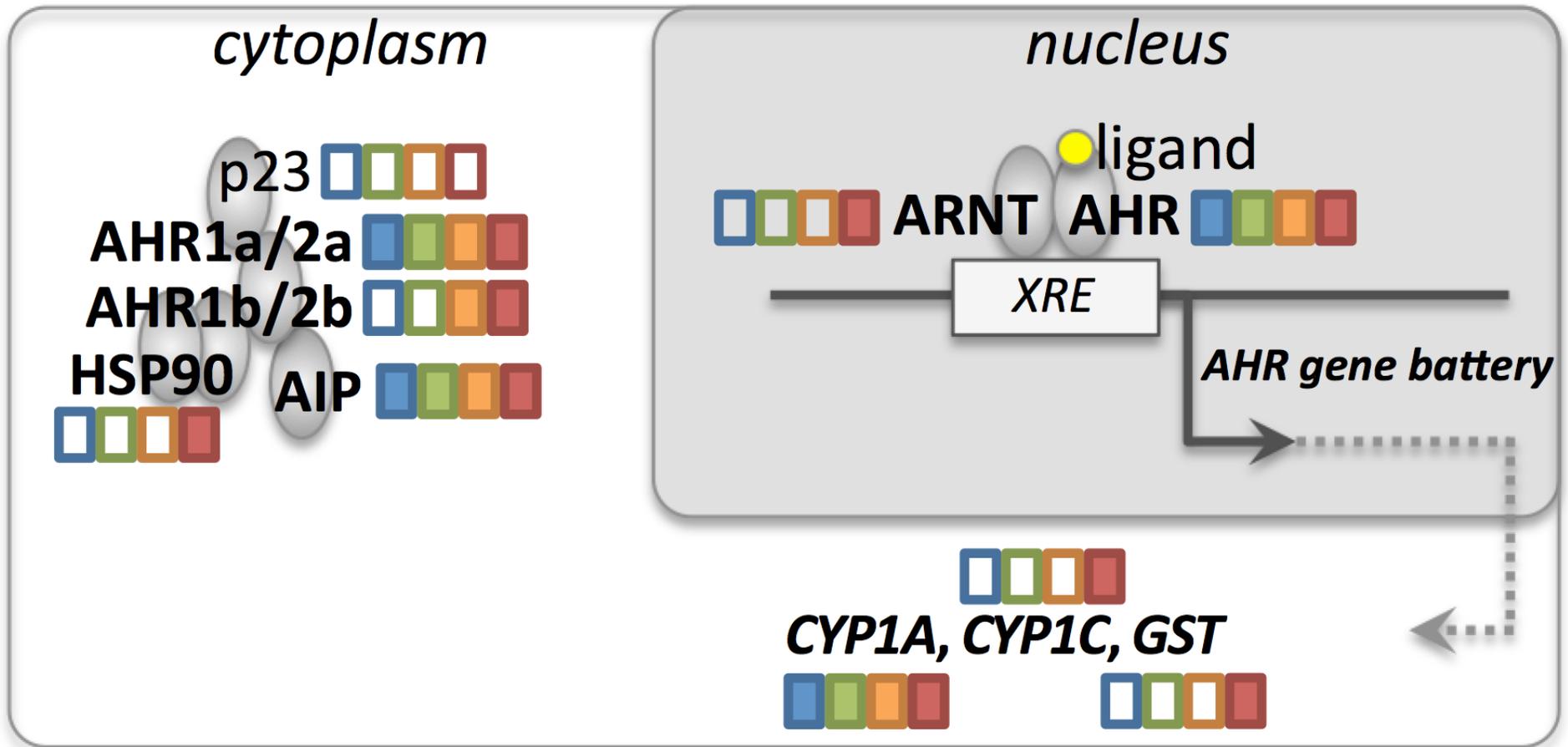


BaP (PAH)



- AHR is a transcription factor (regulates gene expression)
- Activated by TCDD, PCBs, PAHs (and others)
- Involved in mechanism of toxicity in vertebrate animals
- Physiological roles in development and immune system

AHR Pathway as a shared target of selection in multiple populations of resistant killifish



Colors = populations

Filled boxes = under selection

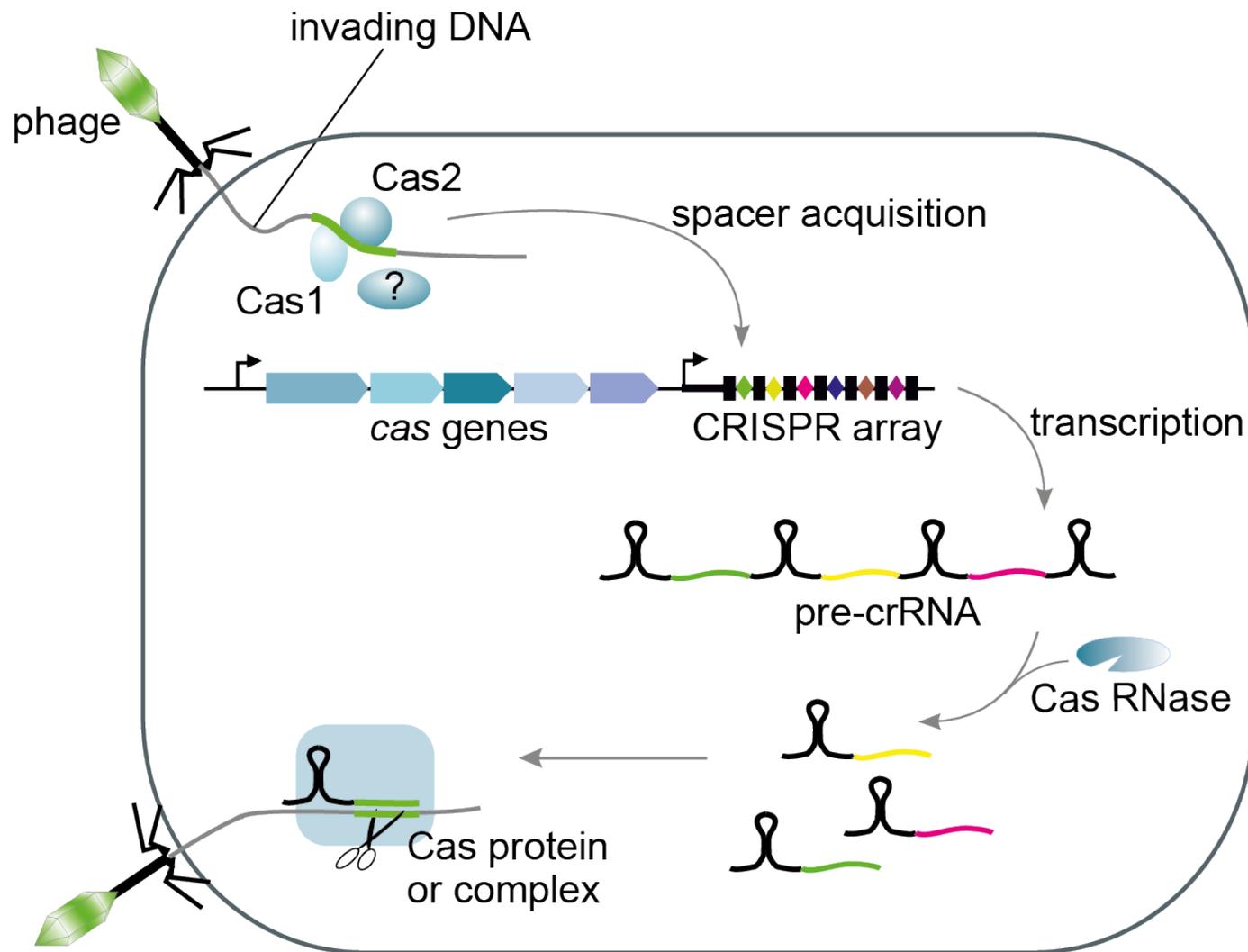
Reid *et al.* 2016 *Science* 354: 1305

Loss-of-function approaches

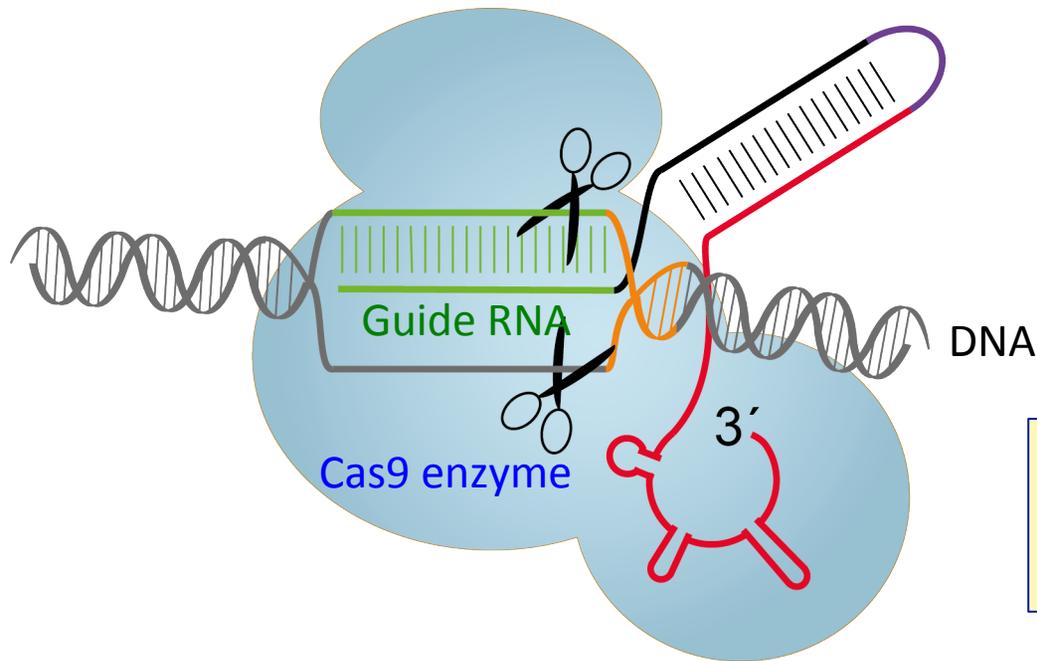
- Widely used way to understand gene function
- Untargeted mutagenesis
 - radiation, chemicals, retroviral
- Knock-down (reduce) expression
 - RNA interference, anti-sense RNA
- Gene knock-out (KO) (homologous recombination)
 - mice (1989)
- Targeted genome editing
 - Zinc finger nuclease (**ZFN**) (2005)
 - Transcription activator-like effector nucleases (**TALENs**)
 - Clustered regularly interspaced short palindromic repeats (**CRISPR**) (2013)

The CRISPR-Cas* adaptive immune system

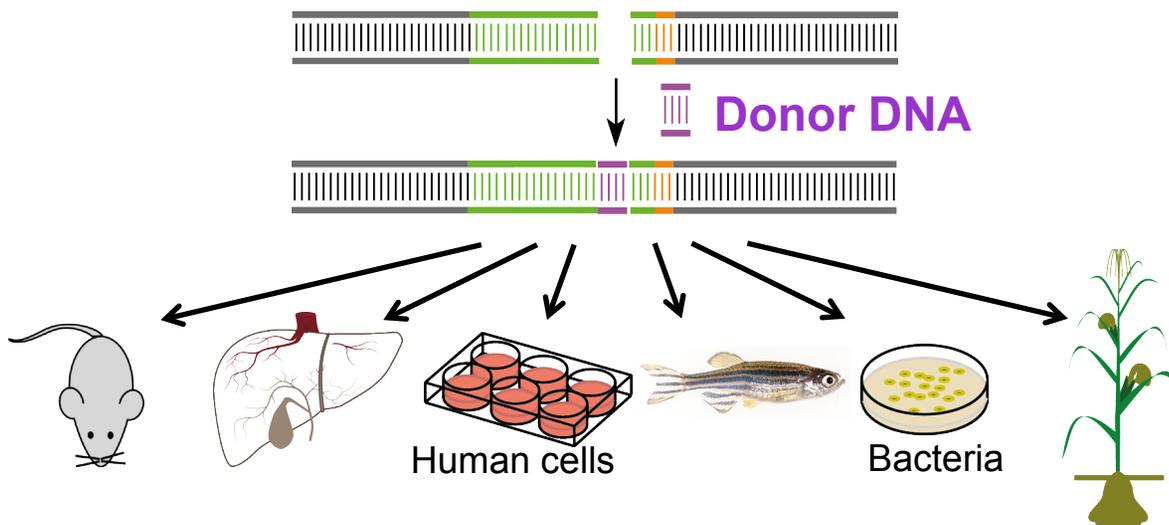
**Clustered Regularly Interspaced Short Palindromic Repeats (CRISPR) and CRISPR-associated (Cas)*



RNA-programmable CRISPR-Cas9

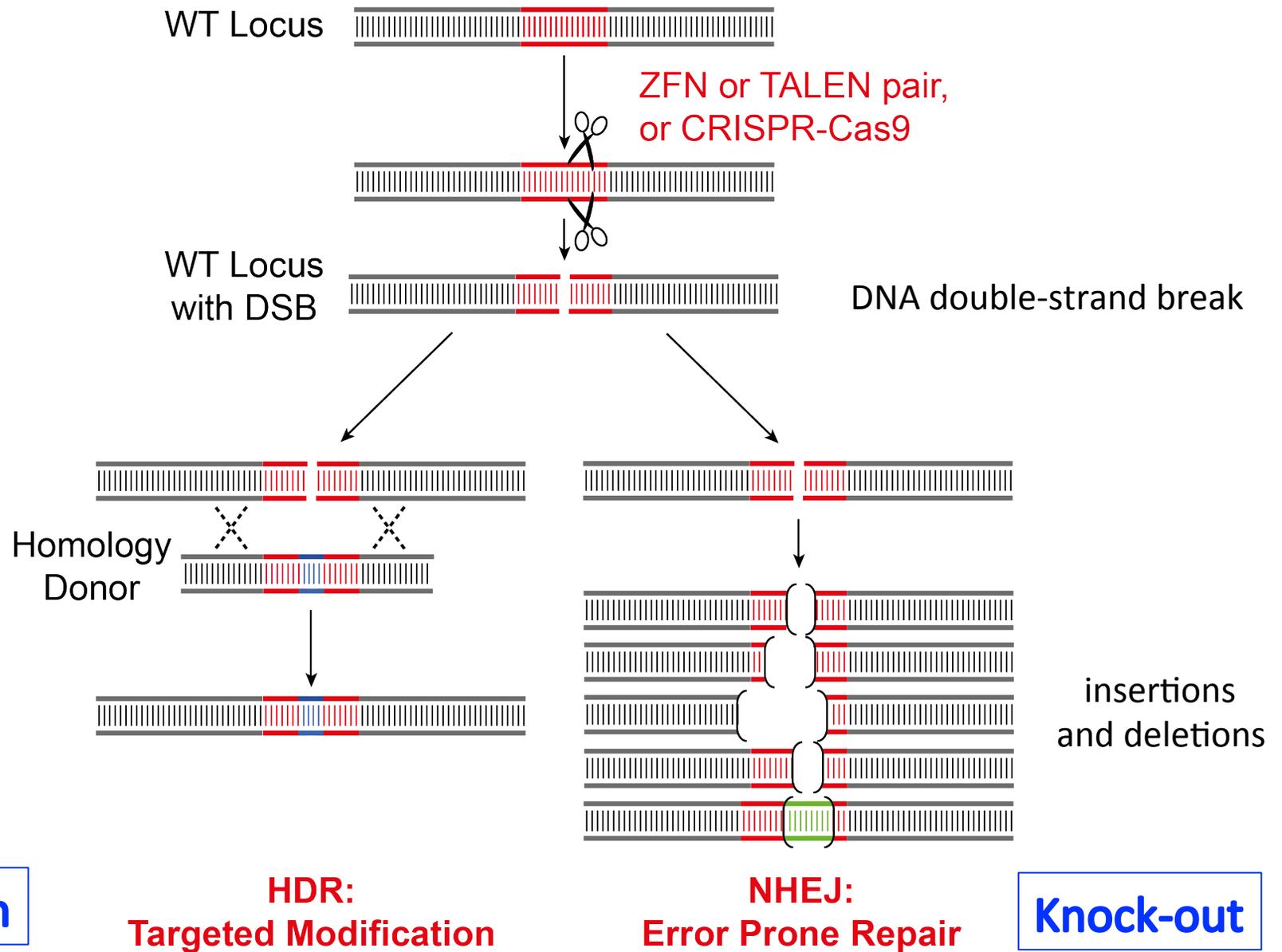


- Guide RNA directs Cas9 to target
- Cas9 endonuclease cuts DNA
- Can be used in most organisms



E. Charpentier (<http://www.nationalacademies.org/gene-editing/Gene-Edit-Summit/index.htm>)

Two types of genome editing with CRISPR-Cas9

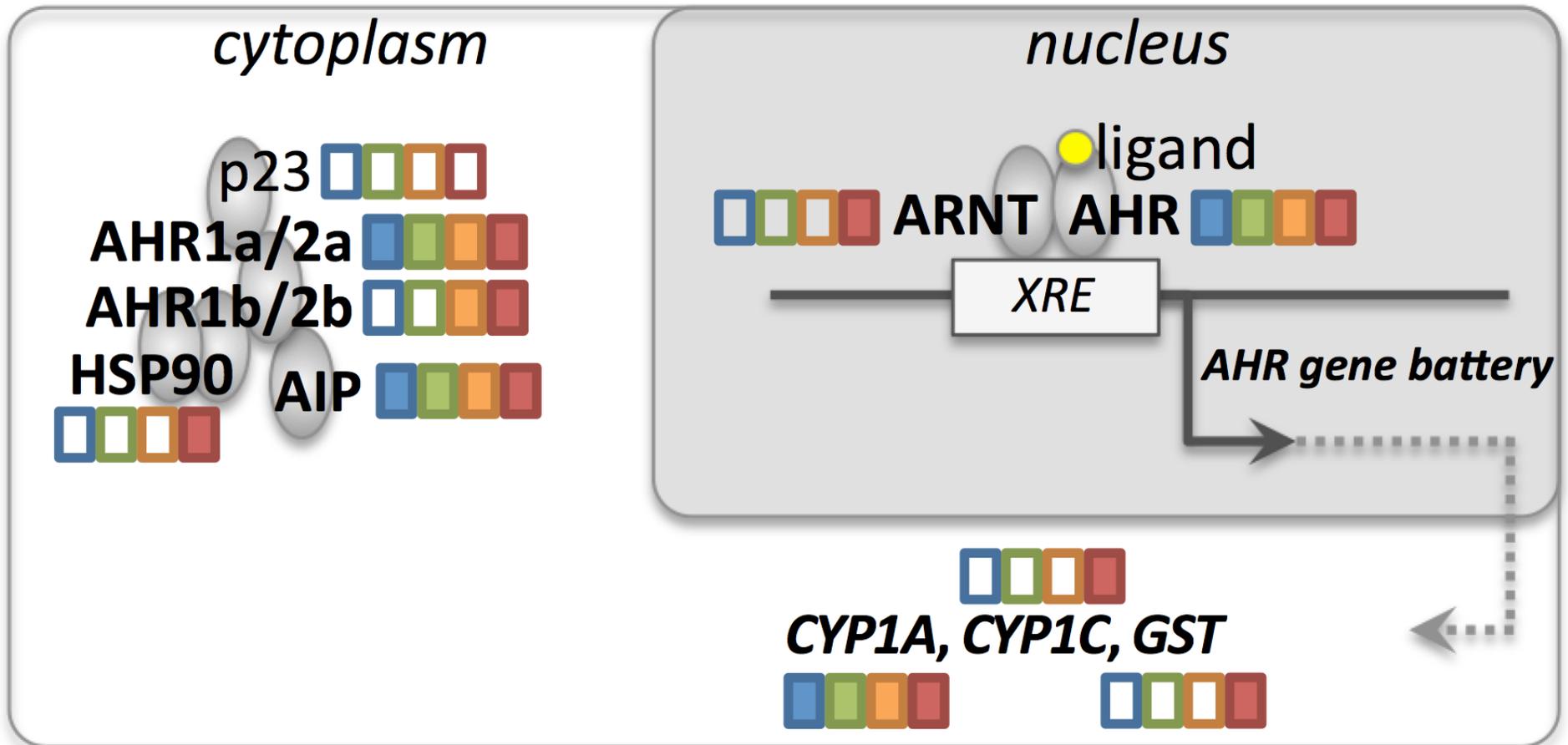


E. Charpentier (<http://www.nationalacademies.org/gene-editing/Gene-Edit-Summit/index.htm>)

Uses of CRISPR-Cas9 (selected examples)

- Targeted gene inactivation (knock-out)
- Precision gene editing, e.g. introduction of SNP
- Genome-wide gene targeting (screening) (Chris Vulpe)
- Tissue-specific gene inactivation (conditional KO)
- Targeted insertion of exogenous gene
- Targeted insertion of fluorescent protein tag (e.g. GFP)
- Targeted tagging of DNA
- Targeted transcriptional regulation (activation or inhibition)
- Targeted epigenetic modification (e.g. methylation)

Using CRISPR to target genes in the AHR Pathway: *AHR1*, *AHR2*, *AIP*



Targeting AHR and AIP genes in zebrafish and killifish

*Raising to adulthood

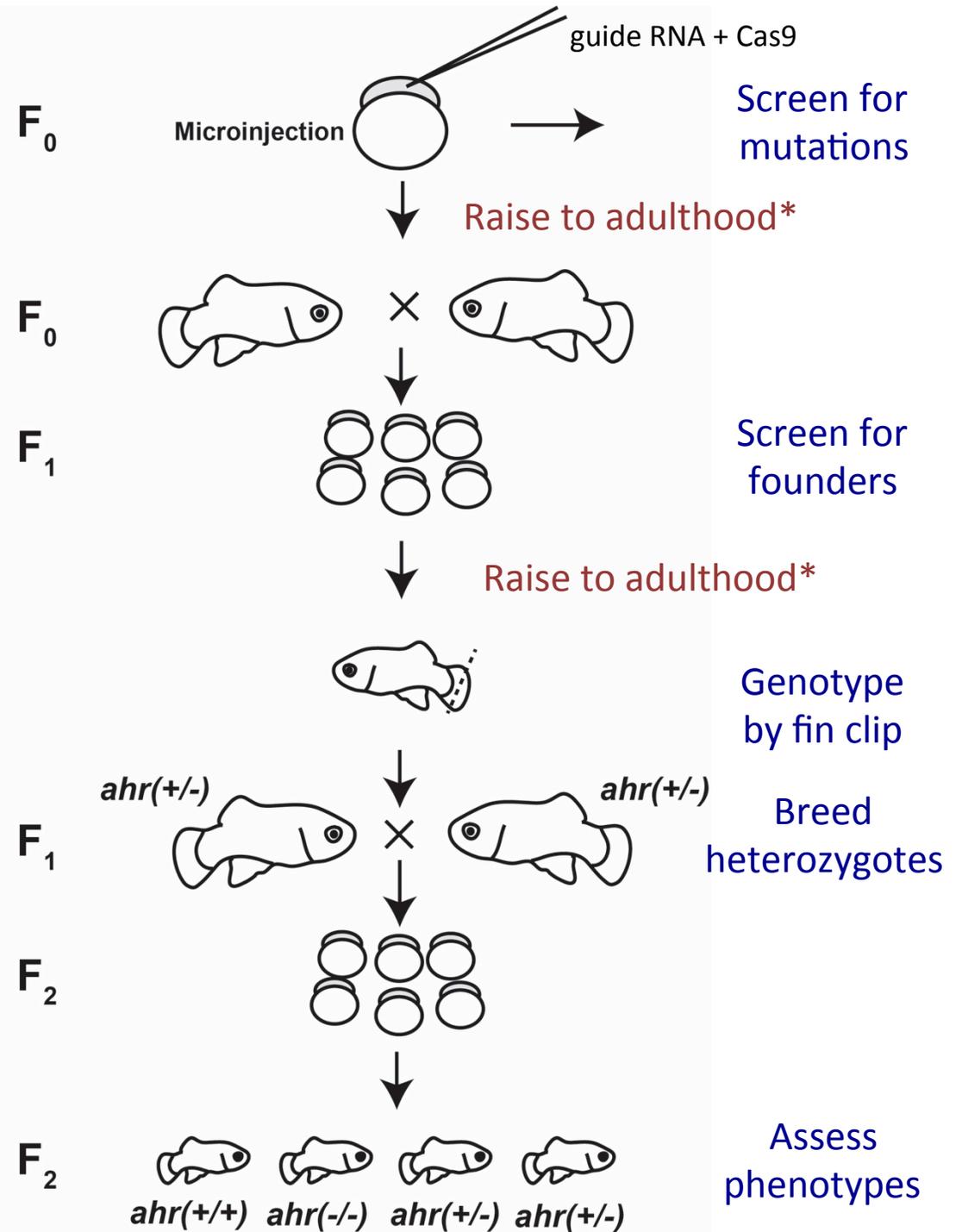


Zebrafish: 3 months



Killifish: 2 years (!)

Aluru *et al.* 2015 *Aquat Toxicol* 158: 192



Progress in generating knock-out and knock-in killifish and zebrafish



Zebrafish:

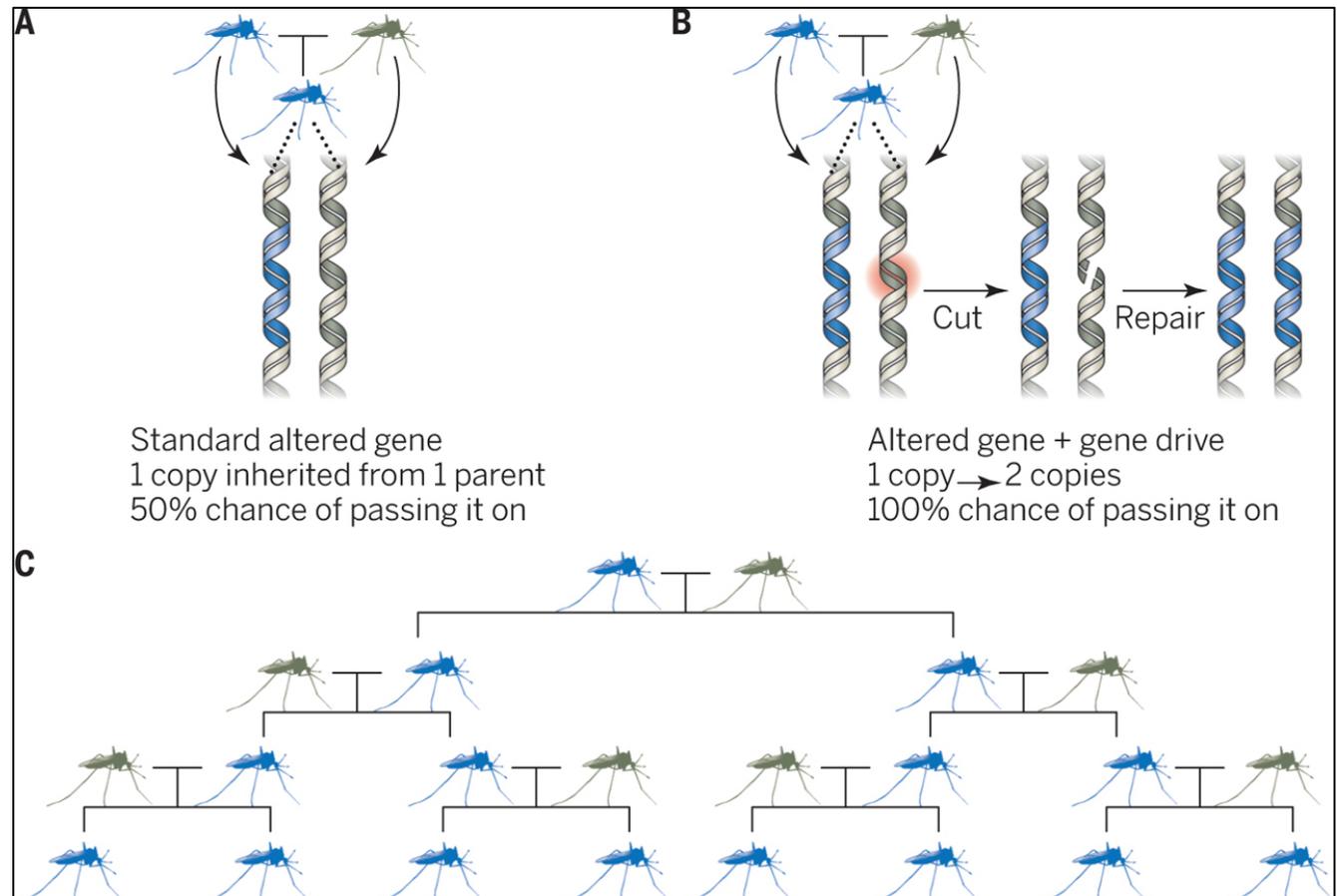
- **AHR1b:** mutant line established
- **AHRRa:** mutant line established
- **AIP mutants:** potential founders being raised and screened
- **AIP knock-ins:** in progress

Killifish:

- **AHR2a:** founders identified
- **AHR2b:** founders identified
- **AIP mutants:** potential founders being raised

We are NOT:

- Releasing genetically engineered fish into the environment.
- Using *Gene Drives* (self-propagating genome editing cassette causes mutagenic chain reaction)



Implications / Significance

- Which genes (AHRs, AIP) control sensitivity to PCBs.
- How long-term exposure leads to functional genetic changes in AHR pathway genes in a population.
- How knock-out (complete loss of function) might differ from knock-down (reduced function), e.g. genetic compensation (basic research).
- Lessons for human health: Humans have homologous genes that control sensitivity to these chemicals.
- What are the costs (trade-offs) of altering the function of these genes?

Thank you!!



Woods Hole
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Noah
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Andrew Whitehead
Noah Reid



Boston University
Superfund Research Program



National Institute of
Environmental Health Sciences

American taxpayers



Additional Information

<http://www.busrp.org>

<http://www.epa.gov/nbh/>

<http://www.whoi.edu/science/B/people/mhahn/hahn.html>

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